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original scientific paper

Characterization of Red Dragon Fruit Wine Fermented with a Newly Identified Yeast Strain *Saccharomyces cerevisiae* M7

Running head: Characterization of Red Dragon Fruit Wine Fermentation

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SUMMARY

Research background. Dragon fruit (*Hylocereus* spp.) has been known to be a rich source of bioactive compounds, such as anthocyanins, betacyanin, betaxanthin and other phenolic substances, and possesses a nutritional profile suitable to produce wine with functional properties. The aim of this study was to characterize the wine fermentation from red dragon fruit juice by a newly identified yeast strain.

Experimental approach. Yeast strains from *banh men*, a Vietnamese traditional alcoholic fermentation starter, were screened for ethanol production using thermally pretreated red

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34 dragon fruit juice. The most potent candidate was identified by the DNA sequencing method
35 and subjected to an optimization study using a one-factor-at-a-time approach to optimize the
36 conditions for red dragon fruit wine fermentation.

37 *Results and conclusions.* Results showed that thermal pretreatment of the red dragon fruit
38 juice at 70 °C for 10 mins resulted in a higher level of phenolic and antioxidants compared
39 with other pretreatment temperatures. Among the 4 isolates, M7 was the most potent alcohol
40 fermenter, which was then identified to be *Saccharomyces cerevisiae* using a DNA
41 sequencing method. The optimal conditions for wine fermentation from red dragon fruit juice
42 by *S. cerevisiae* M7 included a pitching rate of 10⁸ CFU/mL, an initial sucrose content of 18 %
43 m/V, an initial pH of 4.5, fermentation temperature of 30 °C and a fermentation time of 6 days.
44 In such conditions, the wine fermented by *S. cerevisiae* M7 had an ethanol concentration of
45 (12.12±0.15) % V/V total phenolic content of (37.78±0.38) mg GAE/mL, anthocyanin content
46 of (11.22±0.31) mg CGE/L, betacyanin content of (65.18±0.82) mg/L, betaxanthin content of
47 (60.47±1.29) mg/L and antioxidant activity measured by DPPH scavenging capacity of
48 (65.41±0.44) %.

49 *Novelty and scientific contribution.* This study used a novel yeast strain *Saccharomyces*
50 *cerevisiae* M7 for fermentation. In addition, the results of the study present novel data such as
51 the optimal parameters and the accumulation of bioactive compounds
52 (phenolics, anthocyanins, betalains) related to fermentation of red dragon fruit wine.

53

54 **Keywords:** *Saccharomyces cerevisiae*; yeast; wine; red dragon fruit; anthocyanin

55

56 INTRODUCTION

57 Wine is traditionally produced from the fermentation of the grape must. However, the
58 development of wine from fruits other than grapes has gained much interest due to its multiple
59 properties, such as color, flavor, and nutritional values (1). Many fruits from temperate regions
60 (e.g. apples and berries) and tropical regions (e.g. banana, mango, pineapple, and sweet
61 potatoes) can be used to produce wines. As such, wines made from cherries, raspberries, and
62 blueberries contain a significant amount of polyphenols, flavonoids, and anthocyanins that
63 give the wines remarkable antioxidant, anti-proliferative, anti-inflammatory and anti-aging
64 capacities (2–4). Among the fruits from tropical and subtropical regions, dragon fruit
65 (*Hylocereus* spp.) possesses suitable properties to produce wine, evidenced by the pH of the
66 juice ranging from 4.3 to 5 depending on cultivars, while glucose and fructose of the juice
67 being in the ranges of 49.1 to 104 g/L and 19.2 to 29 g/L, respectively (5). The red dragon fruit

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68 is also a rich source of flavonoids, betacyanin and betaxanthin, and carotene (6,7), making
69 wine produced from these fruits a potential functional food.

70 During the production of fermented beverages, the yeast *Saccharomyces cerevisiae*
71 plays an essential role in converting carbohydrates to ethanol and participating in secondary
72 fermentation, which influences flavor and aroma development (8). Therefore, *S. cerevisiae*
73 has been used as a starter culture for various fermented beverages, such as wine, whisky,
74 cognac, sake, and beer (9). In Vietnam, however, *banh men* has been used as a starter to
75 produce traditional alcoholic beverages from rice for centuries. *Banh men* is produced from
76 uncooked rice dough and oriental herbs inoculated with a starter from the previous batch (10).
77 According to Lee and Fujio (11), *banh men* is similar to fermentation starters traditionally used
78 in other Asian countries in terms of microbial composition. Among the microflora present in
79 *banh men*, *S. cerevisiae* strains were reported as the main ethanol producer (10). In this study,
80 we aimed to screen the *S. cerevisiae* strains from *banh men* capable of fermenting red dragon
81 fruit juice into wine and investigate the effect of fermentation conditions on the quality of wine
82 produced by the most potent strain.

83

84 MATERIALS AND METHODS

85 Chemicals

86 All of the reagents used in experiments such as glucose, sucrose, peptone, yeast
87 extract, Hansen broth, Hansen agar, buffer, gallic acid, cyanidin 3-glucoside, Folin-Ciocalteu's
88 phenol, cyanidin-3-glucoside, phenol, chloroform, isoamyl alcohol, ethanol, agarose, Na₂CO₃,
89 KCl, CH₃COONa, 2,2-diphenyl-1-picrylhydrazyl (DPPH) were purchased from Sigma-Aldrich,
90 St. Louis, MO, USA.

91

92 Isolation of yeasts

93 One gram of '*banh men*' was finely ground and suspended in 9 mL of saline. The
94 suspension was further serially diluted with saline and spread onto Hansen agar plates. The
95 plates were then incubated at 28 to 32 °C for 24 to 48 h until obtaining isolated colonies. The
96 colonies were subsequently transferred to the new Hansen agar including 2 % *m/V* of glucose,
97 1 % *m/V* of peptone, 0,1 % *m/V* of yeast extract and 1.5 % *m/V* of agar. The colonies that are
98 round, smooth, and white to whitish cream and the cells that have an oval shape and budding
99 characteristics were suspected to be yeast and used for further studies.

100

101 *Red dragon fruit juice preparation*

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102 The red dragon fruits were washed under running water for about 1 min and drained
103 for 30 min. The fruits were then peeled, and the juice was extracted from the fruit pulp using
104 an electric slow juicer (Hurom H200, Korea). The juice had a total soluble solid of 12.5 ± 0.30
105 % and a pH of (4.53 ± 0.12) . The total phenolic, anthocyanin, betacyanin and betaxanthin
106 contents of the juice were (23.11 ± 0.69) mg GAE/mL, (5.09 ± 0.42) mg CGE/L, (31.21 ± 0.56)
107 mg/L and (23.12 ± 0.39) mg/L, respectively.

108

109 *Thermal treatment of dragon fruit juice and screening of yeast strains*

110 The dragon fruit juice was thermally treated at temperatures 60, 70 and 80 °C for 10
111 min, followed by cooling to ambient temperature. Sucrose was added to the juice samples to
112 obtain a total soluble solid of 18 % *m/V*. The yeast isolates were then inoculated into the juice
113 at 10^7 CFU/mL density and kept at 25 °C for fermentation. When the formation of bubbles
114 ceased, as an indication of primary fermentation completion, the fermented juice samples
115 were analyzed for various quality attributes. The thermal treatment regimen and yeast isolate
116 that yield the highest levels of ethanol, anthocyanins, total phenolic compounds, betacyanin
117 and betaxanthin and antioxidant activity will be selected for further studies.

118

119 *Identification of selected yeast strain*

120 The yeast isolate that produces the wine with higher levels of ethanol and antioxidant
121 substances was identified by Internal Transcribed Spacer Regions (ITS) sequencing method.
122 The ITS sequence was compared against accession numbers available in the GenBank
123 database using the Basic Local Alignment Search Tool (BLAST)(12).

124

125 *Total genomic DNA extraction*

126 The selected yeast isolate was grown in Hansen broth for 24 h, followed by
127 centrifugation at $8000\times g$ (Sigma 1-16; Sigma, Osterode am Harz, Germany) for 2 min to obtain
128 biomass. The biomass was washed with 700 μ L of sterile distilled water. The genomic DNA
129 of the yeast was extracted using the TopPURE® Genomic DNA extraction kit (ABT Biomedical
130 Solutions, HCM city, Vietnam) following the manufacturer's instructions. Briefly, an aliquot of
131 800 μ L lysis buffer was added to the tube containing the biomass, mixed, and added with 40
132 μ L of 20 % *m/V* SDS. The mixture was vortexed for 2 min and incubated at 65 °C for 30 mins,
133 followed by centrifugation at $10\ 000\times g$ for 15 min at 4 °C (Sigma 1-16, Sigma). The collected
134 supernatant was added with an equivalent volume of phenol:chloroform:isoamyl alcohol

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135 (25:24:1), mixed and centrifuged at 10 000×g for 15 min at 4 °C (Sigma 1-16, Sigma,
136 Germany). The top layer was collected, mixed with an equivalent volume of isopropanol, and
137 incubated at – 40 °C for 2 h. The mixture was subsequently centrifuged at 10 000×g for 15
138 min at 4 °C (Sigma 1-16, Sigma, Germany) to collect genomic DNA. The genomic DNA was
139 washed with 500 µL of 70 % *m/V* ethanol twice and resuspended in 30 µL of sterile water. The
140 quality of genomic DNA was verified by electrophoresis on 1 % *m/V* agarose gel. An aliquot
141 of 1 µL of RNase (100 µg/µL) was added to the DNA solution to eliminate RNA.

142

143 *DNA amplification and sequencing analysis*

144 The genomic DNA was amplified by using yeast universal primers ITS1
145 (5'TCCGTAGGTGAACCTGCGG 3') and ITS4 (5'TCCTCCGCTTATTGATATGC 3'). The PCR
146 reaction volume was 60 µL consisting of 30 µL of 2X Go Tag Green Master Mix, 3 µL of each
147 primer (10 pmol/µL), 6 µL of genomic DNA, and 18 µL of H₂O. The PCR was carried out at
148 the following conditions: initial denaturation at 95 °C for 5 min; 30 cycles of denaturation at 95
149 °C for 1 min, annealing at 53 °C for 1 min, and extension at 72 °C for 1 min; and final extension
150 at 72 °C for 10 min. The PCR product was checked by electrophoresis at 70 V for 30 min on
151 1 % agarose gel, stained by SafeView Classic Nucleic Acid Stains (ABM, Inc. USA). The DNA
152 bands on the gel were visualized by using Ultra Slim LED Illuminator and the size of DNA was
153 estimated using GeneRuler 1kb DNA Ladder (Thermal Scientific Inc. USA). The PCR product
154 was sent to 1st BASE (Apical Scientific Sdn. Bhd., Malaysia) for DNA sequencing. The
155 obtained sequences were then aligned and compared with sequences of species available in
156 the NCBI database using BLAST (13).

157

158 *Dragon fruit wine fermentation by selected strain*

159 Effects of fermentation conditions on the quality of fermented dragon juice were
160 evaluated by varying several parameters one by one. The parameters of interest investigated
161 in this study were in the order of pitching rate (10⁵–10⁹ CFU/mL), initial total soluble solid (12–
162 21 °Bx), initial pH (3.5–5.5), fermentation temperature (20–35 °C) and fermentation time (1–7
163 d). The quality of the wine and the efficacy of the yeast strain will be assessed via ethanol
164 concentration, anthocyanins, phenolic content, and antioxidant activity.

165

166 *Wine quality analysis*

167 Ethanol concentration

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168 The ethanol concentration of the fermented juice was determined using the AOAC
169 method 920.57 (14). Briefly, ethanol from 200 mL of the fermented juice was separated by
170 distillation. A hydrometer was used to measure the gravity of the distillates, which were then
171 used to calculate the ethanol concentration.

172

173 Total phenolic content

174 An aliquot of wine was centrifuged at 6000×g for 15 min and used to determine the
175 total phenolic content and antioxidant activity. The total phenolic content was determined
176 according to the method of Singleton and Rossi (15). Briefly, 200 µL of the extract was mixed
177 with 1 mL of 10 % m/V -Folin-Ciocalteu's phenol reagent and 1.2 mL of 10 % m/V -Na₂CO₃
178 solution. The mixture was then allowed to react for 2 h at room temperature and the
179 absorbance was measured at 760 nm. Gallic acid was used as a standard and the total
180 phenolic content was expressed as milligrams of gallic acid equivalent (GAE) per milliliter of
181 the juice (mg/mL).

182

183 Total anthocyanin content

184 The total anthocyanin content was determined by the pH differential method described
185 by Lee *et al.* (16). Briefly, the absorbances of samples diluted in 0.025 M potassium chloride
186 buffer (pH 1) or 0.4 M sodium acetate buffer (pH 4.5) were measured concurrently at 520 and
187 700 nm, respectively, after 20 mins incubation at ambient temperature. The total anthocyanin
188 content was calculated as milligrams of cyanidin 3-glucoside equivalents (CGE) per liter of
189 the juice (mg/L) using the following equation:

$$190 \quad \text{Total anthocyanin content} = \frac{A \cdot M \cdot DF \cdot 10^3}{\epsilon \cdot L} \quad /1/$$

$$191 \quad \text{where } A = (A_{520 \text{ nm}} - A_{700 \text{ nm}})_{\text{pH}=1} - (A_{520 \text{ nm}} - A_{700 \text{ nm}})_{\text{pH}=4.5} \quad /2/$$

192 $A_{520 \text{ nm}}$ is the absorbance of the samples at 520 nm; $A_{700 \text{ nm}}$ is the absorbance of the samples
193 at 700 nm; M is the molecular mass of cyanidin-3-glucoside (449.2 g/mol), DF is the dilution
194 factor, 10^3 is the coefficient for conversion from gram to milligram; ϵ is the molar extinction
195 coefficient of cyanidin-3-glucoside (26 900 L/(mol·cm)) and L is the path length (1 cm).

196

197 Antioxidant activity

198 Antioxidant activity was determined by the DPPH radical scavenging method (17).
199 Briefly, 0.4 mL of the sample or blank (ethanol) was mixed with 3.6 mL of 0.1 mM DPPH. The
200 reaction was allowed to occur in the dark for 1 h at room temperature. The absorbance of the

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201 resulting solution was measured at 517 nm and used to calculate the antioxidant activity of
202 the juice as the DPPH scavenging capacity (%) using the following equation:

$$203 \quad \text{DPPH scavenging capacity} = \frac{A_{\text{Blank}} - A_{\text{Sample}}}{A_{\text{Blank}}} \cdot 100 \quad /3/$$

204 where A_{Blank} is the absorbance of blank and A_{Sample} is the absorbance of the juice extract
205 sample.

206

207 *Betalain content*

208

209 The betalains content was quantified as described previously (18). Briefly, the optical density
210 of the samples was measured at 480 nm and 540 nm for determining betacyanin and
211 betaxanthin contents, respectively. The contents of each betalain compound (mg/mL) were
212 calculated using the following equations:

$$213 \quad \gamma_{\text{bc}} = \frac{A_{540 \text{ nm}} \cdot D_f \cdot M_1 \cdot 1000}{e_1 \cdot L} \quad /4/$$

$$214 \quad \gamma_{\text{bx}} = \frac{A_{480 \text{ nm}} \cdot D_f \cdot M_2 \cdot 1000}{e_2 \cdot L} \quad /5/$$

215 where γ_{bc} and γ_{bx} are the betacyanin and betaxanthin mass content (mg/L), respectively, A_{540}
216 $_{\text{nm}}$ and $A_{480 \text{ nm}}$ are the absorbance of the samples at 540 and 480 nm, respectively, M_1 and M_2
217 are the betaxanthin (308 g/mol) and betacyanin (550 g/mol) molecular mass, respectively, e_1
218 and e_2 are the molar extinction coefficient of betaxanthin (48000 L/mol) and betacyanin (6000
219 L/mol), respectively, D_f is the dilution factor, 1000 is the coefficient for conversion from gram
220 to milligram, and L is the path length (1 cm).

221

222 *Statistical analysis*

223 Data were reported as mean value \pm SD of triplicate experiments. One-way ANOVA,
224 followed by Duncan's test, was used to determine the difference between the means at a
225 significant level of 0.05. Statistical analyses were performed using SPSS V17.0 software (19).

226

227 **RESULTS AND DISCUSSION**

228 *Isolation and screening of yeast strains*

229 Among 32 isolated colonies obtained from Hansen agar plates, 18 isolates (designated
230 as strains M1 to M18) were considered as yeasts based on their cell morphology observed
231 using electron microscopy. These isolates were screened for ethanol production by fermenting

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232 red dragon juice for 24 h at 28 °C. The results showed that four isolates (M2, M7, M11 and
233 M17) produced ethanol (Table S1). These four strains were selected for further experiments.

234 *Effect of thermal pretreatment and yeast isolates on the quality of wine*

235 Temperature is one of the most important factors affecting the extraction of bioactive
236 ingredients from biomass (20). Thus, the thermal pretreatment of dragon fruit juice in this study
237 was expected to affect the quality of the wine in terms of bioactive components, such as
238 phenolics, anthocyanins, betacyanin, betaxanthin and their antioxidant activity.

239 On the other hand, ethanol concentration is another key element determining the yield
240 and activity of bioactive compounds (21). During the fermentation, the yeasts continuously
241 produce ethanol, which consequently alters the ethanol concentration of the fermented juice,
242 hence the extraction of bioactive substances. In this experiment, we study how yeast isolates
243 from *banh men* influence the quality attributes of dragon fruit wine made from juice that was
244 thermally pretreated with different temperatures. The two-way ANOVA showed that thermal
245 pretreatment and yeast isolates and the interaction between these two factors affect the
246 concentrations of ethanol, total phenolic content, anthocyanin content, betacyanin content,
247 betaxanthin content and antioxidant activity of the dragon fruit wine ($p < 0.05$). As such, yeast
248 isolates produced higher levels of ethanol at a pretreatment temperature of 70 °C than at 60
249 or 80 °C (Table 1). Also, the concentrations of phenolic compounds, bioactive compounds
250 (anthocyanins, betacyanin and betaxanthin) as well as antioxidant activity of the wine produced
251 from the juice pretreated at 70 °C were significantly higher than at other temperatures. This
252 result suggested that phenolic compounds were most effectively extracted from dragon fruit
253 pulp particles to the juice at 70 °C. The results from this experiment were in line with a study
254 reported by El Darra *et al.* (22) that thermovinification pretreatment at 70 °C promoted the
255 release of phenolic compounds from grape skin cells. Among the isolates capable of ethanol
256 fermentation, the M7 yielded higher levels of ethanol ($(6.29 \pm 0.33) \% (v/v)$) than M2, M11, and
257 M17 at a pretreatment temperature of 70 °C. In addition, the total phenolic ($(27.22 \pm 5.41) \text{ mg}$
258 GAE/mL), anthocyanins ($(10.02 \pm 0.25) \text{ mg CGE/L}$), Betacyanin ($(50.37 \pm 0.99) \text{ mg/L}$),
259 Betaxanthin ($43.60 \pm 0.72) \text{ mg/L}$) contents, and DPPH scavenging capacity ($57.65 \pm 0.35) \%$ of
260 the juice pretreated at 70 °C and fermented by M7 were markedly greater than by other
261 isolates. In short, results from this study showed that the pretreatment temperature of 70 °C
262 and isolate M7 appeared to be most suitable for producing wine from red dragon fruit juice
263 and were therefore selected for further experiments.

264

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265 *Identification of yeast strain*

266 The BLAST search of the ITS sequence of isolate M7 against reference sequences of
267 *Saccharomyces cerevisiae* species from GenBank (accession numbers of the ITS sequences
268 are KY109257.1 and MZ452353.1) showed a high similarity (>99 %), indicating that isolate
269 M7 belongs to this species (Table 2). Thus, the strain was named *S. cerevisiae* M7.

270

271 *Optimization of red dragon juice fermentation by the yeast strain*

272 Effect of pitching rate

273 Studies showed that pitching rate is a crucial factor affecting the wine fermentation
274 from different fruit juices by yeasts (23,24). In this study, the effect of pitching rate on ethanol
275 production, total phenolic, anthocyanin contents, and antioxidant activities of the dragon fruit
276 wine was investigated. Results show that ethanol and phenolic concentrations, anthocyanin
277 content, betacyanin and betaxanthin contents, and the DPPH scavenging capacity of the
278 fermented juice increased with the increase of pitching rate and reached the maximum levels
279 of (12.5±0.3) % V/V, (35.19±0.90) mg GAE/mL, (9.9±0.20) mg CGE/L, (62.61±0.70) mg/L,
280 (60.51±0.68) mg/L and (63.2±0.3) %, respectively, at the pitching rate of 10⁸ CFU/mL, then
281 leveled off with the further increment of pitching rate (Fig. 1). The increase in ethanol
282 production in response to the increment of inoculation size to a certain level was also observed
283 in a study by Huan *et al.* (25) who reported that the maximum level of ethanol of around 3.5
284 % V/V was obtained at a yeast rate of 2 % V/V after 40–48 h of fermentation, while the lower
285 or higher amount of yeast added to the juice gave lower ethanol productivity. Similarly,
286 Samson *et al.* (26) also documented that ethanol production in pomegranate fruit juice
287 fermentation increases with the increase in the pitching rate from 2 to 8 % V/V, then decreases
288 at higher inoculation sizes. In another study of wine produced from cactus pear and lantana
289 camara fruit juice, a mild level of 10 % V/V of yeast inoculation, compared with 8 or 12 % V/V,
290 was found to be most favorable for wine fermentation (27). In the current study, the
291 improvement of anthocyanin and phenolic compound concentrations and antioxidant activity
292 of the red dragon wine at a higher pitching rate (e.g. 10⁸ CFU/mL) was likely to be yielded by
293 the higher ethanol concentration obtained at this pitching rate, which enhanced the extraction
294 of these substances from the red dragon fruit pulp particles.

295

296 Effect of initial sucrose content

297 As one of the important carbon sources for wine fermentation by yeasts (9), sucrose
298 has been documented as the most influential factor affecting the quality of wine produced from

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299 fruit juice compared with other factors, such as SO₂ treatment, yeast inoculation, and
300 fermentation time (24). In this study, we investigated the fermentation of red dragon fruit wine
301 using *S. cerevisiae* M7 by varying the initial sucrose content from 12 to 21 % *m/V* while the
302 pitching rate and the juice pH were kept constant at 10⁸ CFU/mL and 4.5, respectively. Results
303 showed that higher initial sucrose content led to higher ethanol production and the maximum
304 ethanol yield (12.3±0.33) % *V/V* was obtained at the initial sucrose concentration of 18 % *m/V*
305 (Fig. 2a). Further elevation of initial sugar content did not result in higher ethanol accumulation.
306 Quality attributes, including total phenolic content, anthocyanin content, betacyanin content,
307 betaxanthin content, and antioxidant activity of the wine, were improved with the increase in
308 initial sugar content up to 18 % *m/V* and leveled off beyond this concentration (Fig. 2b, Fig.
309 2c, Fig. 2d, Fig. 2e and Fig. 2f). A maximum level of (36.15±0.43) mg GAE/mL, (10.55±0.37)
310 mg CGE/L, (66.53±0.84) mg/L, (64.31±0.64) mg/L and (64.13±0.88) % was obtained for
311 phenolic, anthocyanin, betacyanin and betaxanthin contents, and antioxidant activity measured
312 by DPPH scavenging capacity, respectively. However, these results were discordant with
313 those reported by Yuan *et al.* (24) that a higher concentration of initial sugar content (*e.g.* 24
314 % *m/V*) is more favorable for ethanol accumulation. In contrast, Arroyo-Lopez *et al.* (28)
315 documented that a sugar concentration higher than 20 % *m/V* decreased the yeast cell growth
316 rate, which may retard the ethanol production rate. In addition, wine fermentation with high
317 sugar concentration negatively affected the production of volatile compounds, which might be
318 detrimental to the quality of the wine (29). The high sucrose content might impose a high
319 osmosis pressure on the yeast cells, reducing their fermentation performance. It was
320 concluded from this study that an initial sucrose concentration of 18 % *m/V* –was the most
321 favorable to obtain red dragon fruit wine with a high antioxidant profile.

322

323 Effect of initial pH

324 Environmental pH is another critical factor for the growth of yeast and wine
325 fermentation (28), as it has been known to alter the conformation, hence the function of cellular
326 membrane-embedded proteins and eventually affect the fermentation rate and constitutions
327 of fermentation products (30). To investigate the effect of initial pH on the fermentation of red
328 dragon fruit wine by *S. cerevisiae* M7, the pH of the juice was adjusted to 3.5, 4.5, and 5.5
329 before yeast inoculation. Results showed that ethanol productivity was ~ 12 % *V/V* at the initial
330 pH levels of 3.5 and 4.5 (Fig. 3a). The total phenolic compounds were ~ 37 mg GAE/mL at
331 initial pH 3.5 (Fig. 3b). Anthocyanin content was slightly higher at initial pH of 3.5 than at 4.5
332 (Fig. 3c) while there was no significant difference in betacyanin and betaxanthin contents at

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333 these initial pH (Fig. 3d and Fig. 3e). The antioxidant activity was ~65 % at the initial pH 3.5
334 (Fig. 3f). Increasing the initial pH of the juice to 5.5 substantially decreased ethanol production
335 as well as the phenolic, anthocyanin, betalainins contents and antioxidant activity of the wine
336 (Fig. 3). The results from our study agreed with those from Liu *et al.* (30) reporting that an
337 initial pH of 4.5 is most favorable for the growth and alcoholic fermentation of several *S.*
338 *cerevisiae* strains, including Freddo, BH8 and N°.7303. It must be noted, however, optimal pH
339 for wine fermentation might be strain dependent as *S. cerevisiae* NCIM 3045 give the highest
340 yield of ethanol at a pH of 5.5 in the palm wine fermentation (31). Nonetheless, results from
341 the current study indicated that the initial pH of 4.5 was optimal for wine fermentation from red
342 dragon fruit juice by *S. cerevisiae* M7 regarding the ethanol productivity, phenolic, betacyanin,
343 betaxantin contents and antioxidant activity of the wine.

344

345 Effect of temperature

346 Culture temperature is an important factor affecting the physiology of microorganisms
347 during fermentation. Consequently, the accumulation of fermented products in the culture is
348 impacted (26). In this work, the strain *S. cerevisiae* M7 was used for the fermentation of red
349 dragon fruit juice at various temperatures, ranging from 20 to 35 °C, with pitching rate of 10⁸
350 CFU/mL, 18 % *m/V* of sucrose concentration supplement in red dragon fruit juice, and an initial
351 pH of 4.5. The results (Fig. 4) showed the highest content of alcohol (13.1±0.24) % *V/V*, total
352 phenolic compounds ((37.58±0.87) GAE/mL), anthocyanin ((11.41±0.38) CGE/L), betacyanin
353 ((65.63±0.49) mg/L) and betaxantin ((61.15±0.95) mg/L) at a fermentation temperature of 30
354 °C (Fig. 4a, Fig. 4b, Fig. 4c, Fig. 4d and Fig. 4e). There was no significant difference in
355 antioxidant activities at 25 °C and 30 °C (Fig. 4f). The concentration of all these compounds
356 in culture was significantly lower at fermentation temperatures of 20 and 35 °C. These results
357 are similar with those of Patil *et al.* (32) who reported an optimal temperature of 27.5 °C for
358 fermentation of sugarcane–papaya juice by *S. cerevisiae* (EC1118), while the optimal
359 temperatures in fermentation of palm juice (31) and pomegranate juice (26) were reported to
360 be higher, at 32 °C and 37 °C, respectively.

361

362 Effect of fermentation time

363 Fermentation time is critical to determine the quality of wine as well as to obtain a wine
364 with a high level of bioactive compounds. In this study, the changes in ethanol production,
365 phenolic compounds, and antioxidant of red dragon wine were investigated during the primary

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366 fermentation period. As shown in Fig. 5a, the ethanol concentration of wine gradually
367 increased to a maximum level of $(12.12 \pm 0.15) \% V/V$ from the first to the sixth day of
368 fermentation, while further fermentation beyond this time did not yield any additional amount
369 of ethanol. In parallel with the increase in ethanol concentration, the levels of phenolic content,
370 anthocyanins, betacyanin, betaxanthin contents and antioxidant activity of the wine also
371 steadily increased during the first six days of fermentation and reached a maximum level of
372 $(37.78 \pm 0.38) \text{ mg GAE/mL}$, $(11.22 \pm 0.31) \text{ mg CGE/L}$, $(65.18 \pm 0.82) \text{ mg/L}$, $(60.47 \pm 1.29) \text{ mg/L}$
373 and $(65.41 \pm 0.44) \%$, respectively (Fig. 5b, Fig. 5c, Fig. 5d, Fig. 5e and Fig. 5f). These results
374 indicated that the primary fermentation period of six days was optimal for *S. cerevisiae* M7 to
375 produce red dragon fruit wine. In other studies, Samson *et al.* (26) and Yuan *et al.* (24)
376 reported that the fermentation time of 7 or 8 days is optimal for wine fermentation from
377 pomegranate or green jujube juices, respectively. The variation in the optimal time for wine
378 fermentation between studies might be attributed to the differences in yeast strains used,
379 inoculation size, fermentation temperature, and the nature of juices as these parameters
380 greatly affect the growth and fermentation ability of yeast strains.

381

382 CONCLUSION

383 Our result demonstrate that the phenolic, anthocyanin, betacyanin, betaxanthin content
384 and antioxidant ability of the wine product produced from the red dragon fruit juice pretreated
385 at 70 °C significantly increased. Among the yeast isolates tested, *Saccharomyces cerevisiae*
386 M7 exhibited the highest levels of these compounds in wine production. The optimal
387 fermentation parameters for this strain was established. They are a pitching rate of 10^8
388 CFU/mL, an initial sugar content of 18 % *m/V*, an initial pH of 4.5, a temperature of 30 °C, and
389 a fermentation time of 6 days. Under these conditions, the levels of ethanol, total phenolics,
390 anthocyanins, betacyanins, betaxanthins, and antioxidant activity in this wine product were
391 remarkably high. These findings highlight the potential of red dragon fruit juice as a substrate
392 for producing high-quality wine with enhanced bioactive properties using the novel strain
393 *Saccharomyces cerevisiae* M7 as a starter.

394

395 CONFLICT OF INTEREST

396 The authors declare that they have no conflict of interest.

397

398 AUTHORS' CONTRIBUTION

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399 T. T. Q. Anh contributed to the design of the study, data collection, and drafting of the
400 article. N.T. An contributed to data analysis, interpretation, and drafting of the article. D. T. B.
401 Thuy contributed to overall project management, study design, and drafting of the article. All
402 authors contributed to the final approval of the version to be published.

403

404 SUPPLEMENTARY MATERIAL

405 Supplementary materials are available at: www.ftb.com.hr.

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411

412 REFERENCES

413 1. Jagtap UB, Bapat VA. Wines from fruits other than grapes: Current status and future
414 prospectus. Food Biosci. 2015;9:80–96.

415 <https://doi.org/10.1016/j.fbio.2014.12.002>

416 2. Jeong JH, Jung H, Lee SR, Lee HJ, Hwang KT, Kim TY. Anti-oxidant, anti-proliferative
417 and anti-inflammatory activities of the extracts from black raspberry fruits and wine. Food
418 Chem. 2010;123(2):338–44.

419 <https://doi.org/10.1016/j.foodchem.2010.04.040>

420 3. Su MS, Chien PJ. Antioxidant activity, anthocyanins, and phenolics of rabbiteye blueberry
421 (*Vaccinium ashei*) fluid products as affected by fermentation. Food Chem. 2007;104(1):182–
422 7.

423 4. Sun SY, Jiang WG, Zhao YP. Evaluation of different *Saccharomyces cerevisiae* strains on
424 the profile of volatile compounds and polyphenols in cherry wines. Food Chem.
425 2011;127(2):547–55.

426 <https://doi.org/10.1016/j.foodchem.2011.01.039>

427 5. Esquivel P, Stintzing FC, Carle R. Comparison of morphological and chemical fruit traits
428 from different pitaya genotypes (*Hylocereus* sp.) grown in Costa Rica. J Appl Bot Food Qual.
429 2007;81(1):7–14.

430 6. Le Bellec F, Vaillant F, Imbert E. Pitahaya (*Hylocereus* spp.): A new fruit crop, a market
431 with a future. Fruits. 2006;61(4):237–50.

432 7. Wu LC, Hsu HW, Chen YC, Chiu CC, Lin YI, Ho JAA. Antioxidant and antiproliferative

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- 433 activities of red pitaya. Food Chem. 2006;95(2):319–27.
- 434 8. Walker GM, Stewart GG. *Saccharomyces cerevisiae* in the production of fermented
435 beverages. Beverages. 2016;2(4):1–13.
- 436 9. Berry DR, Slaughter JC. Alcoholic Beverage Fermentations. In: Lea AGH, Piggott JR,
437 editors. Fermented Beverage Production. Boston, MA: Springer US; 2003. p. 25–39.
438 https://doi.org/10.1007/978-1-4615-0187-9_2
- 439 10. Thanh VN, Thuy NT, Chi NT, Hien DD, Ha BTV, Luong DT, *et al.* New insight into
440 microbial diversity and functions in traditional Vietnamese alcoholic fermentation. Int J Food
441 Microbiol. 2016;232:15–21.
442 <https://doi.org/10.1016/j.ijfoodmicro.2016.05.024>
- 443 11. Lee AC, Fujio Y. Microflora of banh men, a fermentation starter from Vietnam. World J
444 Microbiol Biotechnol. 1999;15(1):57–62.
- 445 12. Leaw SN, Chang HC, Sun HF, Barton R, Bouchara JP, Chang TC. Identification of
446 medically important yeast species by sequence analysis of the internal transcribed spacer
447 regions. J Clin Microbiol. 2006;44(3):693–9.
- 448 13. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL.
449 BLAST+: Architecture and applications. BMC Bioinformatics. 2009;10:421.
450 <https://doi.org/10.1186/1471-2105-10-421>
- 451 14. AOAC Official Method 920.57. Alcohol in wines by volume from specific gravity.
452 Rockville, MD, USA: AOAC International; 1990.
- 453 15. Singleton VL, Rossi JJA. Colorimetry of total phenolics with phosphomolybdic-
454 phosphotungstic acid reagents. Am J Enol Vitic. 1965;16(3):144–58.
455 <https://www.ajevonline.org/cgi/content/abstract/16/3/144>
- 456 16. Lee J, Durst RW, Wrolstad RE. Determination of total monomeric anthocyanin pigment
457 content of fruit juices, beverages, natural colorants, and wines by the pH differential method:
458 Collaborative study. J AOAC Int. 2005;88(5):1269–78.
- 459 17. Perumal S, Klaus B. Antioxidant properties of various solvent extracts of total phenolic
460 constituents from three different agroclimatic origins of drumstick tree (*Moringa oleifera*
461 Lam.) Leaves. J Agric Food Chem. 2003;51(8):2144–55.
- 462 18. Li X, Zhang ZH, Qiao J, Qu W, Wang MS, Gao X, *et al.* Improvement of betalains
463 stability extracted from red dragon fruit peel by ultrasound-assisted microencapsulation with
464 maltodextrin. Ultrason Sonochem. 2022;82:105897.
465 <https://doi.org/10.1016/j.ultsonch.2021.105897>
- 466 19. SPSS Statistics, v. 17.0, SPSS Inc, Chicago, IL, USA; 2008. Available from:

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- 467 <https://www.ibm.com/products/spss-statistics>
- 468 20. Gan CY, Latiff A a. Optimisation of the solvent extraction of bioactive compounds from
469 *Parkia speciosa* pod using response surface methodology. Food Chem. 2011;124(3):1277–
470 83. <https://linkinghub.elsevier.com/retrieve/pii/S0308814610009271>
- 471 21. Ho UTN, Tran LTM, Dinh AQ, Nguyen AT. Response Surface Optimization of Ethanolic
472 Extraction of Antioxidants from Artichoke Leaves. J Food Process Preserv.
473 2015;39(6):1036–44.
- 474 22. El Darra N, Turk MF, Ducasse MA, Grimi N, Maroun RG, Louka N, *et al.* Changes in
475 polyphenol profiles and color composition of freshly fermented model wine due to pulsed
476 electric field, enzymes and thermovinification pretreatments. Food Chem.
477 2016;194(2016):944–50. <https://doi.org/10.1016/j.foodchem.2015.08.059>
- 478 23. Pereira AP, Mendes-Ferreira A, Oliveira JM, Estevinho LM, Mendes-Faia A. High-cell-
479 density fermentation of *Saccharomyces cerevisiae* for the optimisation of mead production.
480 Food Microbiol. 2013;33(1):114–23. <https://doi.org/10.1016/j.fm.2012.09.006>
- 481 24. Yuan L, Li G, Yan N, Wu J, Due J. Optimization of fermentation conditions for fermented
482 green jujube wine and its quality analysis during winemaking. J Food Sci Technol.
483 2022;59(1):288–99. <https://doi.org/10.1007/s13197-021-05013-8>
- 484 25. Huan PT, Hien NM, Anh NHT. Optimization of alcoholic fermentation of dragon fruit juice
485 using response surface methodology. Food Res. 2020;4(5):1529–36.
- 486 26. Samson, Singh AK, Singh G. Optimum parameters for wine production from
487 pomegranate fruit juice. Int J Pharm Sci Res. 2017;8(11):1000–6.
- 488 27. Tsegay ZT. Total titratable acidity and organic acids of wines produced from cactus pear
489 (*Opuntia-ficus-indica*) fruit and Lantana camara (*L. Camara*) fruit blended fermentation
490 process employed response surface optimization. Food Sci Nutr. 2020;8(8):4449–62.
- 491 28. Arroyo-López FN, Orlić S, Querol A, Barrio E. Effects of temperature, pH and sugar
492 concentration on the growth parameters of *Saccharomyces cerevisiae*, *S. kudriavzevii* and
493 their interspecific hybrid. Int J Food Microbiol. 2009;131(2–3):120–7.
- 494 29. Lu Y, Chan LJ, Li X, Liu SQ. Effects of sugar concentration on mango wine composition
495 fermented by *Saccharomyces cerevisiae* MERIT.ferm. Int J Food Sci Technol.
496 2018;53(1):199–208. <https://doi.org/10.1111/ijfs.13574>
- 497 30. Liu X, Jia B, Sun X, Ai J, Wang L, Wang C, *et al.* Effect of Initial PH on Growth
498 Characteristics and Fermentation Properties of *Saccharomyces cerevisiae*. J Food Sci.
499 2015;80(4):M800–8. <https://doi.org/10.1111/1750-3841.12813>
- 500 31. Ghosh S, Chakraborty R, Raychaudhuri U. Optimizing process conditions for palm

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- 501 (*Borassus flabellifer*) wine fermentation using response surface methodology. Int Food Res
502 J. 2012;19(4):1633–9.
- 503 32. Patil PS, Deshannavar UB, Ramasamy M, Emani S. Production, optimization, and
504 characterization of sugarcane (*Saccharum officinarum*)–papaya (*Carica papaya*) wine using
505 *Saccharomyces cerevisiae*. Environ Technol Innov. 2021;21:101290.
- 506
- 507

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508 **TABLES AND FIGURES**509 **Table S1.** Screening of yeast strains for ethanol production

Yeast strain No.	Ethanol concentration/%	Yeast strain No.	Ethanol concentration/%
M1	-	M10	-
M2	3.97	M11	5.12
M3	-	M12	-
M4	-	M13	-
M5	-	M14	-
M6	-	M15	-
M7	5.16	M16	-
M8	-	M17	4.35
M9	-	M18	-

510 “-” represents no ethanol production

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512 **Table 1.** Effect of thermal pretreatment and yeast isolates on the quality of dragon fruit wine

Pretreatment temperature/°C	Yeast isolate	Ethanol concentration/%	Total phenolic content as w(GAE)/(mg/mL)	Anthocyanin content as w(CGE)/(mg/L)	Antioxidant activity (% DPPH scavenged)	γ (betacyanin)/(mg/L)	γ (betaxanthin)/(mg/L)
60	M2	(3.97±0.13) ^{ef}	(14.80±2.10) ^{cde}	(3.60±0.45) ^g	(38.12±0.46) ^{ef}	(26.87±1.14) ^f	(26.41±0.60) ^g
	M7	(5.16±0.38) ^{bcd}	(13.39±1.28) ^{def}	(6.71±0.38) ^{cde}	(42.43±0.60) ^c	(46.63±0.92) ^b	(38.47±0.65) ^c
	M11	(5.12±0.39) ^{bcd}	(18.95±2.16) ^{bcd}	(5.64±0.46) ^{ef}	(39.37±0.65) ^{de}	(44.67±0.93) ^b	(35.53±0.84) ^{de}
	M17	(4.35±0.24) ^{def}	(12.10±1.80) ^{ef}	(5.92±0.39) ^{def}	(28.12±0.11) ⁱ	(35.40±0.75) ^{de}	(30.62±1.00) ^f
70	M2	(4.70±0.29) ^{cde}	(22.71±1.14) ^{ab}	(4.93±0.54) ^f	(46.11±0.42) ^b	(26.64±0.97) ^f	(28.37±0.73) ^{fg}
	M7	(6.29±0.33) ^a	(27.22±5.41) ^a	(10.02±0.25) ^a	(57.65±0.35) ^a	(50.37±0.99) ^a	(43.60±0.72) ^a
	M11	(5.55±0.20) ^{abc}	(20.28±1.63) ^{bc}	(7.457±0.40) ^{bc}	(40.42±0.67) ^d	(46.15±0.48) ^b	(39.13±0.88) ^b
	M17	(5.00±0.19) ^{bcd}	(20.57±1.16) ^{bc}	(6.86±0.35) ^{cde}	(35.58±0.59) ^{fg}	(44.49±0.77) ^b	(36.50±1.23) ^{cde}
80	M2	(3.97±0.23) ^{ef}	(16.50±1.41) ^{bcde}	(5.01±0.40) ^f	(30.66±1.04) ^h	(28.29±0.94) ^f	(28.29±0.94) ^{fg}
	M7	(5.72±0.41) ^{ab}	(15.63±1.20) ^{cde}	(8.62±0.55) ^b	(42.76±0.70) ^c	(41.41±0.49) ^c	(41.41±0.49) ^{ab}
	M11	(4.69±0.43) ^{cde}	(11.85±1.24) ^{ef}	(6.93±0.32) ^{cd}	(35.28±0.33) ^g	(37.70±0.58) ^d	(37.70±0.58) ^{cd}
	M17	(3.65±0.16) ^f	(8.91±1.21) ^f	(6.51±0.53) ^{cde}	(24.25±0.58) ^j	(34.97±0.69) ^e	(34.97±0.69) ^e

513 CGE=cyanidin 3-glucoside equivalent, GAE=gallic acid equivalent. Results are mean values of triplicate analyses±standard deviation. Mean
 514 values in the same column without a common letter differ significantly, p≤0.05
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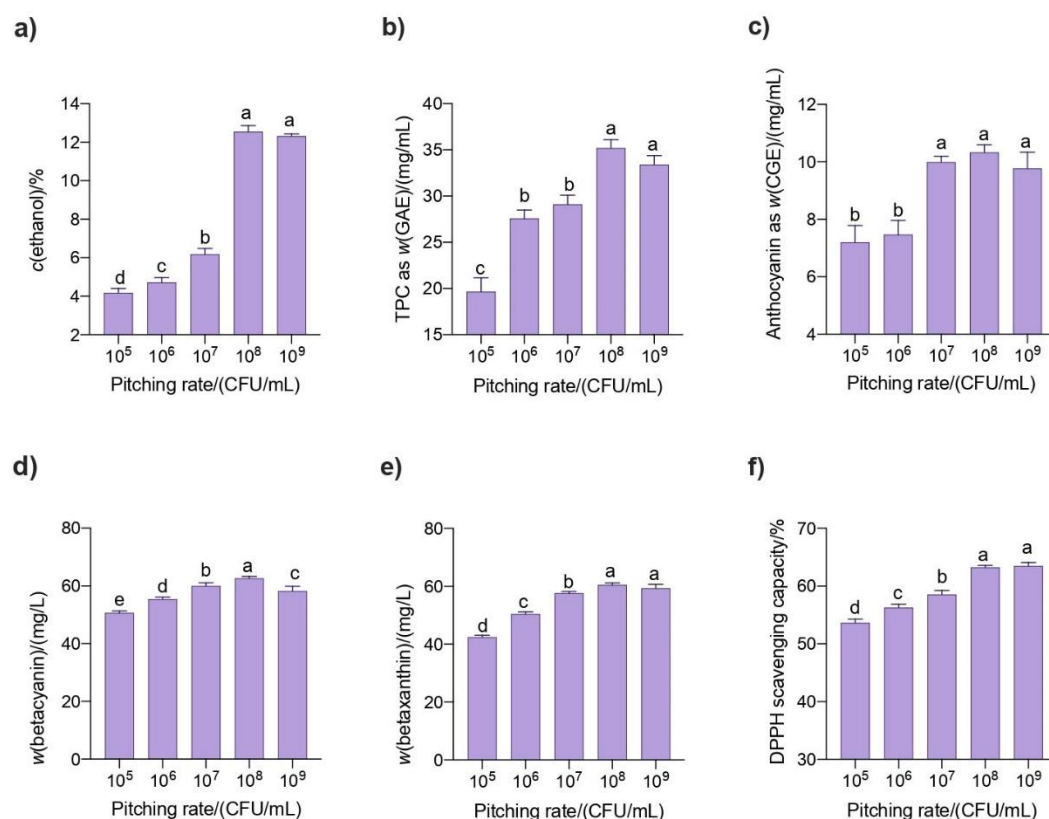
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520 **Table 2.** Identification result of M7 strain

Yeast strain No.	Accession numbers of the ITS sequences	Species	Similarity (%)
M7	KY109257.1	<i>Saccharomyces cerevisiae</i>	100
	MZ452353.1	<i>Saccharomyces cerevisiae</i>	99

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523

524 **Fig. 1.** Effect of pitching rate on: a) ethanol concentration, b) total phenolic content, c) total
 525 anthocyanin content, d) betacyanin content, e) betaxanthin content, and f) antioxidant activity
 526 of the red dragon fruit wine. The juice, with a total soluble solid content of 18% w/v and a pH
 527 of 4.5, was fermented by *S. cerevisiae* M7 at different pitching rates for 7 days at 25 °C. Data
 528 are means of triplicate analyses ± standard deviation. Mean values in each graph without a
 529 common letter differ significantly (p ≤ 0.05)

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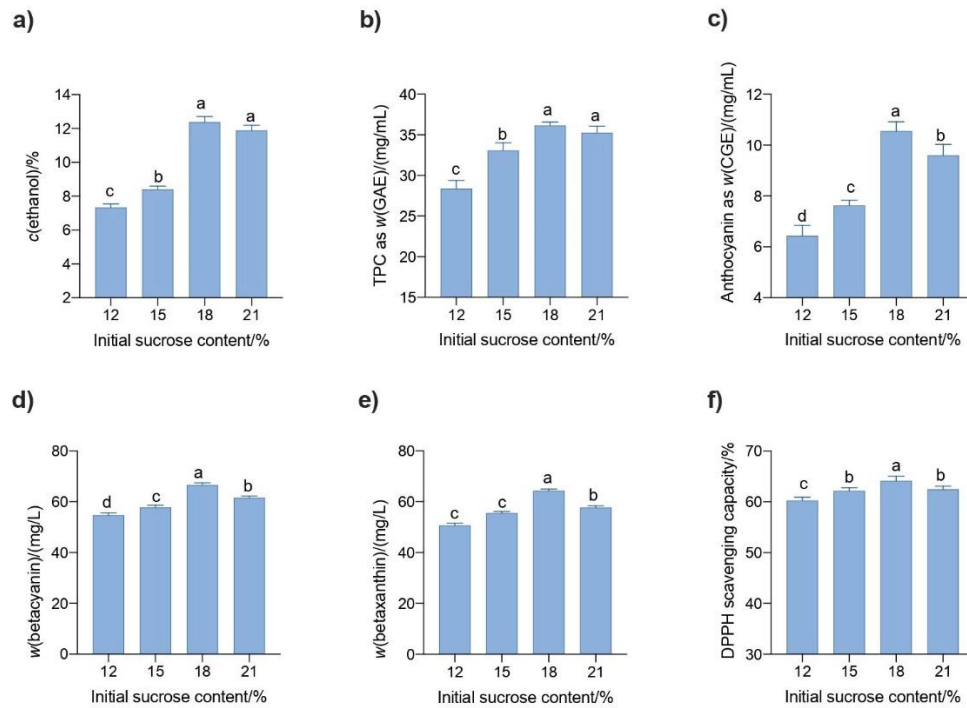
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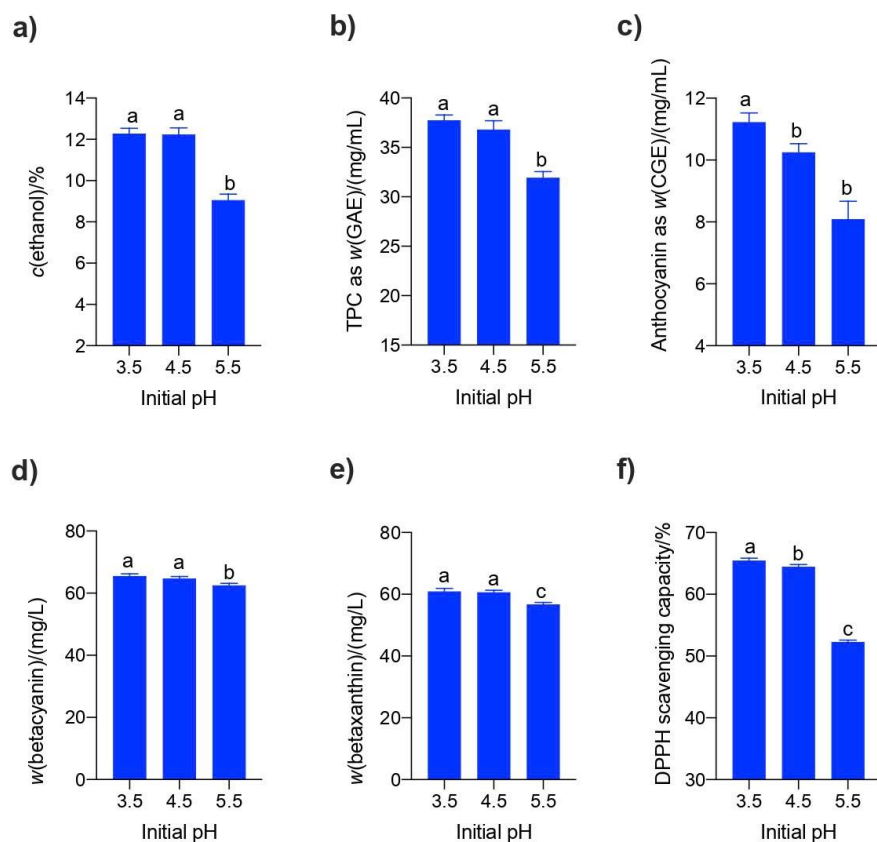


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542 **Fig. 2.** Effect of initial sugar content on: a) ethanol concentration, b) total phenolic content, c)
 543 total anthocyanin content, d) betacyanin content, e) betaxanthin content, and f) antioxidant
 544 activity of the red dragon fruit wine. The juice with various initial sugar content and a pH of 4.5
 545 was fermented by *S. cerevisiae* M7 at a pitching rate of 10^8 CFU/mL at 25 °C for 7 days. Data
 546 are means of triplicate analyses \pm standard deviations. Means in each graph without a
 547 common letter differ significantly, $p \leq 0.05$

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549

550 **Fig. 3.** Effect of pH on: a) ethanol concentration, b) total phenolic, c) total anthocyanin, d)
551 betacyanin, e) betaxanthin contents, and f) antioxidant activity of the red dragon fruit wine.
552 The juice with various initial pH values and a sugar content of 18 % w/v was fermented by *S.*
553 *cerevisiae* M7 at a pitching rate of 10^8 CFU/mL at 25 °C for 7 days. Data are means of triplicate
554 analyses \pm standard deviations. Mean values in each graph without a common letter differ
555 significantly, $p \leq 0.05$

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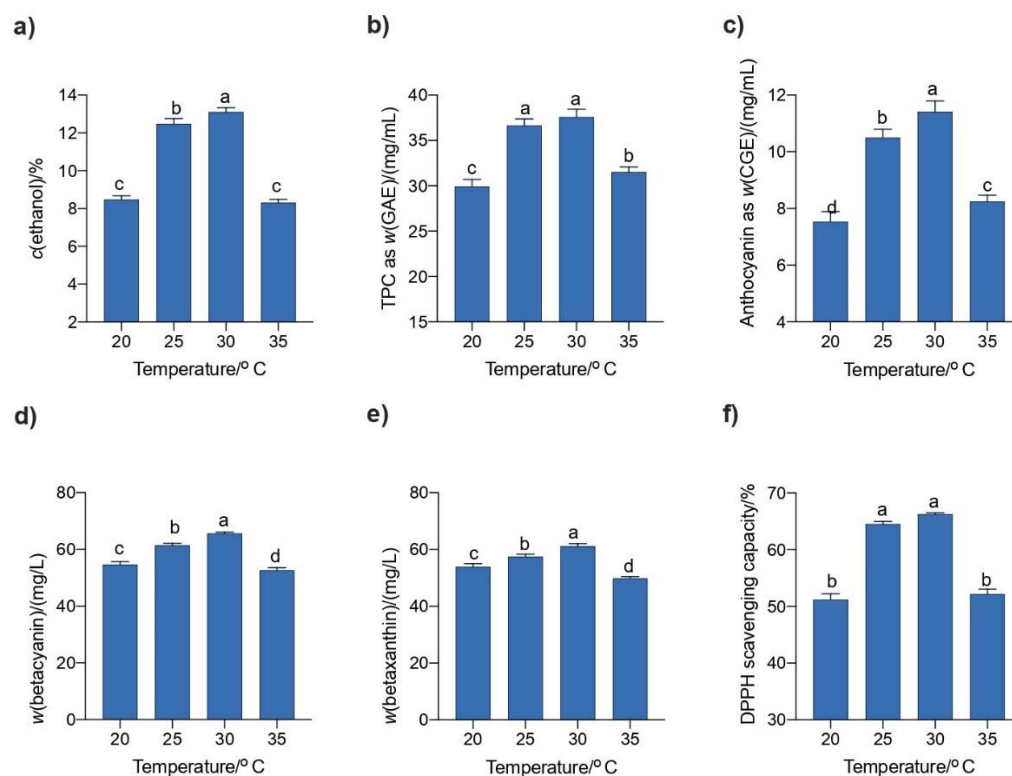
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570 **Fig. 4.** Effect of fermentation temperature on: a) ethanol concentration, b) total phenolic
 571 content, c) total anthocyanin content, d) betacyanin content, e) betaxanthin content, and f)
 572 antioxidant activity of the red dragon fruit wine. The fermentation temperature was varied from
 573 20 to 35 °C. The initial pH was 4.5, sugar content was 18 % w/v, and *S. cerevisiae* M7 was
 574 inoculated at a pitching rate of 10^8 CFU/mL with a fermentation time of 7 days. Data are means
 575 of triplicate analyses \pm standard deviations. Means values in each graph without a common
 576 letter differ significantly ($p \leq 0.05$)

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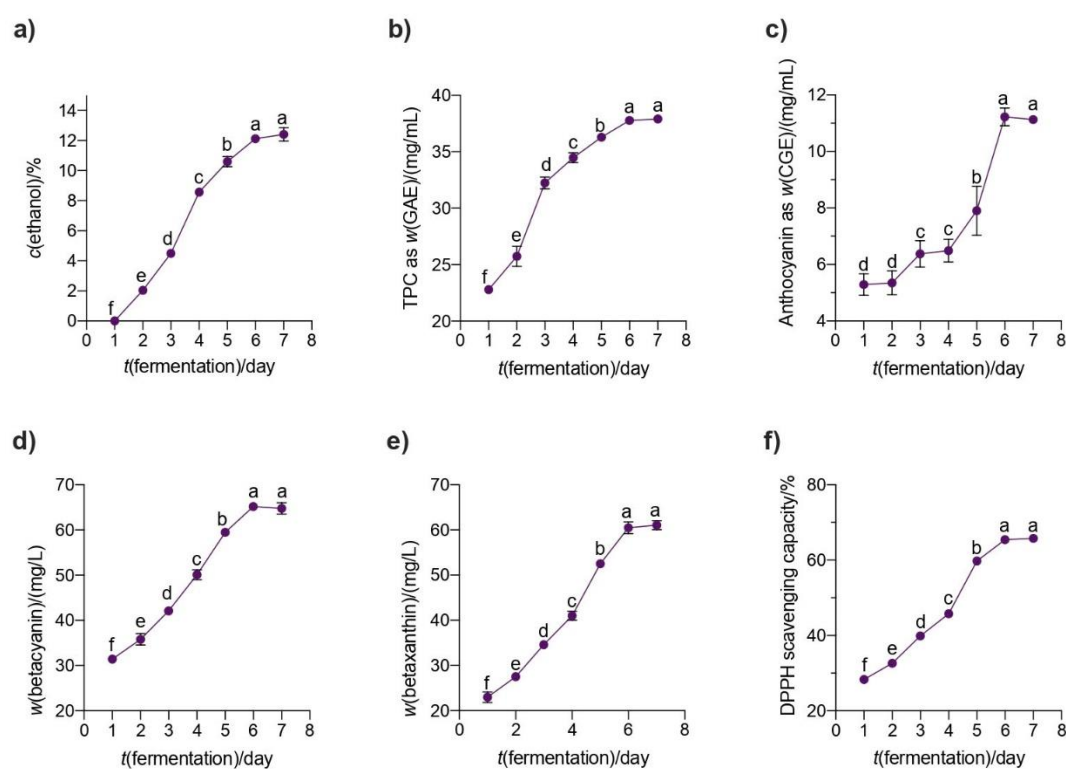
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591 **Fig. 5.** Changes in: a) ethanol concentration, b) total phenolic content, c) total anthocyanin
592 content, d) betacyanin content, e) betaxanthin content, and f) antioxidant activity of the red
593 dragon fruit wine during fermentation. The juice with an initial pH of 4.5 and a sugar content
594 of 18% w/v was fermented by *S. cerevisiae* M7 at 30 °C and a pitching rate of 10⁸ CFU/mL for
595 different times (day). Data are means of triplicate analyses ± standard deviations. Means
596 values in each graph without a common letter differ significantly, p ≤ 0.05